Phylogenetic study of the section *Adonanthe* of genus *Adonis* L. (Ranunculaceae) based on ITS sequences

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ABSTRACT: DNA sequences of nrITS regions obtained from 49 accessions representing 12 species and one variety within the section *Adonanthe* of the genus *Adonis* were analyzed to test the previous intra-sectional classification system and to determine their phylogenetic relationships. The results showed that the seven accessions of *A. amurensis* Regel et Radde included in the present study did not form a monophyletic group, as some of the accessions showed a close relationship with the Japanese endemic species *A. ramosa* Franch., which implies that the current species delimitation and identification of *A. amurensis* is problematic. *Adonis pseudoamurensis* W. T. Wang, which is frequently misidentified as *A. ramosa* in Korea, formed a segregated group, which suggests that they should not be considered as conspecific taxa. Accessions from *A. shikokuensis* Nishikawa et Koji Ito, *A. multiflora* Nishikawa et Koji Ito, and *A. pseudoamurensis* formed a clade, but monophyly of each species was not evident. The nrITS data did not support the classification system proposed by Wang, who classified sect. *Adonanthe* into four series, as most of these were found to be either polyphyletic or paraphyletic.

Keywords: *Adonis*, *Adonanthe*, phylogenetic analysis, sequence variation, ITS

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The genus *Adonis* L. belongs to the family Ranunculaceae and is composed of perennial and annual herbaceous plants that are included in the tribe Adonideae under the subfamily Ranunculoideae (Nishikawa and Kadota, 2006; Ren et al., 2009). Approximately 26–30 species grow in the northern temperate zone including Asia, Europe, and North America, and some annual plants are known to be distributed from Southwest Asia to North Africa, as well as along the shores of the Mediterranean (Meusel et al., 1965; Cronquist, 1981; Mabberley, 1990).

After the initial attempt by Linnaeus (1753), De Candolle (1818) established the classification system, including the genus *Adonis* at the section level, in which 10 taxa of this genus were divided into the sect. *Adonia* DC., an annual species with 5–10 petals, 18–20 stamens, erect styles, and cylindrical or globular aggregated achenes, and the sect. *Consiligo* DC., a perennial species with 8–15 petals, 25–30 stamens, recurved styles, and globular aggregated achenes. There have been various arguments on the classification of the sect. *Consiligo* DC. For instance, unlike the sect. *Adonia* DC., this group was subdivided into subsection levels (Bobrov, 1937; Poschkurlat, 1977; Tamura, 1990, 1991) and some taxa were treated as an independent genus (gen. *Adonanthe* Spach) or section (sect. *Ancistrocarpium* Spach) (Spach, 1839).

Through a morphological study on 30 taxa of the genus *Adonis* L. distributed globally that represented a comprehensive study of this genus, Wang (1994a, 1994b) promoted sect. *Adonia* (annual plants) and sect. *Consiligo* DC. (perennial plants) to the subgenus *Adonis* (DC.) W. T. Wang and subgen. *Adonanthe* (Spach) W. T. Wang, respectively. Furthermore it was suggested that the subgen. *Adonis* should be further divided into 3 sections and 2 series, while the subgen. *Adonanthe* should be divided into 3 sections and 4 series, resulting in a new classification system with 1 genus, 2 subgenera, 6 sections, and 6 series.

The sect. *Adonanthe* W. T. Wang in the subgen. *Adonanthe* is divided into the series *Amurense* Poschk., which is characterized by ovate, ovate deltoid, or elliptic shaped leaves and yellow or white petals; ser. *Coeruleae* Poschk. with oblong or ovate oblong shaped leaves and white or purple petals; ser. *Apeninae* Bobr. ex Poschk. with 3-pinnately compound leaves; and ser. *Vernales* Bobr. ex Poschk. with 3-pinnatisect palmately compound leaves. Son and Ko (2013) have supported this classification system within the sect. *Adonanthe* based on the morphology of aggregated achenes and the microstructure of the achene surface in the genus *Adonis* L. found in East Asia; however, most of the previous studies on the genus *Adonis* L. mentioned above focused on the classification system based on morphological differences, including those associated with leaf shape and achene characters, whereas a phylogenetic relationship-based classification system of the genus *Adonis* L. has not yet been established. In addition, Son (2015) indicated that, although leaf shape and leaf type in the genus *Adonis* L. could be used for identification in some species, they were highly variable among individuals and in most cases generally did not provide any information for phylogenetic analysis.

Because plants of the sect. *Adonanthe* have similar external morphological characteristics that are highly variable and objective taxonomic characteristics of the taxa within the genus have not been established, it is difficult to identify taxa and investigate their phylogenetic relationships. *A. davidii* Franch. is a Chinese endemic species described by Franchet (1885), but it was considered a synonym of *A. brevistyla* Franch. (Wang, 1980). Later, it was reestablished as an independent taxon by Wang (1994a), whereas *A. brevistyla* was treated as a synonym of *A. davidii* by Fu and Robinson (2001), which shows the different opinions about the taxonomic status of these taxa.

*A. pseudoamurensis* W. T. Wang (Korean name: Gae-bok-su-cho) was introduced as a new species by Wang (1980) based on samples collected in Jilin, China (C. Shen 1204, PE), but it was later considered a synonym of *A. ramosa* Franch. (Korean name: Ga-ji-bok-su-cho) along with *A. multiflora* Nishikawa et Koji Ito (Korean name: Se-bok-su-cho) by the same author (Wang, 1994a). However, Nishikawa and Kadota (2006) regarded *A. ramosa* as a Japanese endemic species and Wang’s (1980) *A. pseudoamurensis* as a synonym of *A. multiflora*. In contrast, Lee et al. (2003) and Son and Ko (2012) recognized *A. pseudoamurensis*, *A. multiflora*, and *A. ramosa* as independent species. In addition, *A. pseudoamurensis* was treated as a variety of *A. amurenensis* [var. *pseudoamurensis* (W. T. Wang) Y. N. Lee] (Lee, 2004, 2006). Thus diverse opinions have appeared about its taxonomic status.

*A. coerulea* Maxim. was described by Maximowicz (1877) based on samples collected from Gansu, China (*N. M. Pczewalski s.n.*, E). The species is characterized by plants glabrous; leaves oblong, 2–3 pinnately compound, with long petioles, densely arranged at the base of the stem; and petals purple. Based on the samples collected from Xizang, China (Z. Qing et al. 76-10534, PE), Wang (1980) designated the population with short pubescence on the stem and leaves, unlike glabrous *f. coerulea*, as *f. puberula* W. T. Wang, and later Wang (1994b) elevated it to var. *puberula* (W. T. Wang) W. T. Wang. However, Fu and Robinson (2001) considered var. *puberula* a synonym of the basic species and recorded it
as a hairless plant except for pistils, adding further to the uncertainty of the taxonomic identity of the f. puberula population, which has been recognized for having short pubescence on the stem and leaves.

Recently, molecular data have been frequently used to establish a phylogenetic classification system. In particular, the ITS regions, which are nuclear DNA regions characterized by patterns of parental inheritance, evolve more rapidly than the coding regions, leading to higher levels of variation among closely related individuals. Thus, the ITS regions have been utilized to investigate inter-species and inter-genus relationships, as well as evolutionary trends and patterns in genetic variation (Baldwin, 1992; Álvarez and Wendel, 2003).

Previous studies on the genus *Adonis* L. in East Asia include sequence analyses of trnL-F and the ITS region sequenced from Japanese *Adonis* plants (Kaneko et al., 2008) as well as an ITS sequence analysis from Korean *Adonis* plants (Suh et al., 2002). Additionally, despite the taxonomic applications of various DNA sequence analyses, which have been completed by a number of taxonomists, the genus *Adonis* L. has only been used as a control for studying relationships with other taxa within the family (Hoot, 1995; Sun et al., 2001; Després et al., 2003; Cai et al., 2009). Furthermore, no representatives from the entire taxa of the sect. *Adonanthe* under gen. *Adonis* L. in East Asia have been phylogenetically classified.

Thus, in the present study, we investigated the phylogenetic relationships of the sect. *Adonanthe* of genus *Adonis* L. using sequences from the ITS region of nuclear ribosomal DNA and aimed to evaluate the validity of the intra-genus classification system more objectively. In addition, we also attempted to address the taxa with species delimitation problems.

**Materials and Methods**

**Materials**

Plant samples from 11 taxa with 21 representative accessions from the sect. *Adonanthe* of genus *Adonis* were either collected directly from the field by the participants in this study or obtained from specimens that were deposited in herbaria (HNHM, PE, and KUN) in Korea and China. Nuclear acid sequence information for *A. vernalis* L. and *A. autumnalis* Nishikawa et Koji Ito were retrieved from GenBank. Data from GenBank were also added for the taxa with different taxonomic status among the sect. *Adonanthe*. *A. annua* L. belongs to the sect. *Adonis* of subgen. *Adonis*, and *Trollius acaulis* Lindl., *T. asiaticus* L., and *T. chinensis* Bunge were selected as the outgroup (Johansson, 1999). The collection data regarding the 13 taxa with 49 accessions and 4 taxa as the outgroup with 4 accessions that were used in the present study are summarized in Table 1.

**DNA extraction, DNA amplification, and sequencing**

Leaf samples collected from specimens or the field were stored in silica gel until completely dehydrated and crushed using a grinder (Mixer Mill MM200, Retsch, Haan, Germany) or a mortar, and total genomic DNA was extracted using a DNA extraction kit (MG10 Plant Genomic DNA Extraction SV mini, Macrogen, Seoul, Korea). Extracted DNA was assessed by electrophoresis using a 0.9% agarose gel. The reaction mixture utilized to amplify DNA using polymerase chain reaction (PCR) included 5 μL 10× Pfu DNA buffer, 1 μL 10 mM dNTP, 1 μL 10 pmol primers in each direction, 25–100 ng template DNA, 0.5 μL Pfu DNA polymerase, and 41.5 μL D.W. in a total volume of 50 μL. The primers used for DNA amplification were ITS4 and ITS5 (White et al., 1990). PCR conditions for gene amplification were as follows: the reaction began with a pre-denaturation step at 94°C for 5 min, followed by 35 thermal cycles of denaturation at 94°C for 30 sec, annealing at 52°C for 1.5 min, and extension at 72°C for 1.5 min, followed by a final extension at 72°C for 7 min. The PCR product was purified with a commercial kit (PCR Purification Kit, Macrogen, Korea), followed by cycle sequencing using a BigDye® terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, USA). Sequences were analyzed using the ABI PRISM 3730XL Analyzer (Applied Biosystems, USA). Sequences were deposited to GenBank and accession numbers were obtained (KU570388–KU570408).

**Alignment of DNA sequences and analyses of phylogeny**

DNA sequences for each sample in both the forward and reverse orientations were assembled and aligned using Geneious ver. 7.1.7 (Biomatters, Ltd., Auckland, New Zealand), and final alignments were manually verified. Maximum Parsimony analysis (MP) and Maximum Likelihood (ML) were conducted using PAUP* ver. 4.0b10 (Swofford, 2003). Heuristic search was carried out with ACCTRAN, MULPARS, and TBR branch swapping options turned on. To determine the support levels for the resulting cladograms, the bootstrap values (Felsentein, 1985) and posterior probabilities (PP) from the Bayesian inference were determined. Bootstrap values were obtained from 1,000 bootstrap replicates. To calculate the posterior probabilities using a Bayesian inference, a substitution model best fit to each set of sequence data was selected based on Akaike Information Criterion (AIC; Posada and Buckley, 2004) using Modeltest ver. 3.7 (Posada and Buckley, 2004).
Table 1. List of taxa used for the ITS analysis with vouchers and Genbank accession numbers.

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Abbr.</th>
<th>Locality and Voucher</th>
<th>Ribotype</th>
<th>Accession No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. davidii Franch.</td>
<td>DAV1</td>
<td>Yunnan, China 02577 (PE)</td>
<td>Z2(27NS, 2ID)</td>
<td>KU570394</td>
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<tr>
<td>A. brevistyla Franch.</td>
<td>BRE1</td>
<td>?, China</td>
<td>Z(26NS, 3ID)</td>
<td>KF233849*</td>
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<tr>
<td>A. brevistyla Franch.</td>
<td>BRE2</td>
<td>Shaanxi, China 2 (KUN)</td>
<td>Z1(27NS, 2ID)</td>
<td>KU570392</td>
</tr>
<tr>
<td>A. sutchuenensis Franch.</td>
<td>SUT1</td>
<td>Sichuan, China 70817 (PE)</td>
<td>T(19NS, 2ID)</td>
<td>KU570406</td>
</tr>
<tr>
<td>A. sutchuenensis Franch.</td>
<td>SUT2</td>
<td>Hubei, China 22832 (PE)</td>
<td>U(19NS, 2ID)</td>
<td>KU570407</td>
</tr>
<tr>
<td>A. sutchuenensis Franch.</td>
<td>SUT3</td>
<td>Gansu, China 1383 (PE)</td>
<td>V(20NS, 2ID)</td>
<td>KU570408</td>
</tr>
<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU1</td>
<td>Mt. Unmu, Gangwon, Korea s.n. (HNHM)</td>
<td>D(9NS)</td>
<td>KU570388</td>
</tr>
<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU2</td>
<td>Mt. Odae, Gangwon, Korea s.n. (HNHM)</td>
<td>D(9NS)</td>
<td>KU570389</td>
</tr>
<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU3</td>
<td>Daekwanryung, Gangwon, Korea</td>
<td>E(10NS, 2ID)</td>
<td>AF454929*</td>
</tr>
<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU4</td>
<td>Mt. Choomguryng, Gyeonggi, Korea</td>
<td>F(10NS)</td>
<td>AF454928*</td>
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<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU5</td>
<td>Mt. Jeoksang, Jeonbuk, Korea</td>
<td>E(10NS, 2ID)</td>
<td>AF454927*</td>
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<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU6</td>
<td>Nemuro, Hokkaido, Japan</td>
<td>A</td>
<td>AB361619*</td>
</tr>
<tr>
<td>A. amurensis Regel et Radde</td>
<td>AMU7</td>
<td>Shari, Hokkaido, Japan</td>
<td>A</td>
<td>AB361620*</td>
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<tr>
<td>A. ramosa Franch.</td>
<td>RAM1</td>
<td>Fujimi, Nagano, Japan s.n. (HNHM)</td>
<td>A</td>
<td>KU570403</td>
</tr>
<tr>
<td>A. ramosa Franch.</td>
<td>RAM2</td>
<td>Hokkaido, Japan, Komineyama-4 (PE)</td>
<td>C(4NS)</td>
<td>KU570404</td>
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<tr>
<td>A. ramosa Franch.</td>
<td>RAM3</td>
<td>Fujisato, Akita, Japan</td>
<td>A</td>
<td>AB361613*</td>
</tr>
<tr>
<td>A. ramosa Franch.</td>
<td>RAM4</td>
<td>Hachinohe, Aomori, Japan</td>
<td>A</td>
<td>AB361612*</td>
</tr>
<tr>
<td>A. ramosa Franch.</td>
<td>RAM5</td>
<td>Tatami, Fukushima, Japan</td>
<td>A</td>
<td>AB361615*</td>
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<tr>
<td>A. ramosa Franch.</td>
<td>RAM6</td>
<td>Asahi, Gifu, Japan</td>
<td>B(1ID)</td>
<td>AB361617*</td>
</tr>
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<td>A. ramosa Franch.</td>
<td>RAM7</td>
<td>Urakawa, Hokkaido, Japan</td>
<td>A</td>
<td>AB361610*</td>
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<tr>
<td>A. ramosa Franch.</td>
<td>RAM8</td>
<td>Furano, Hokkaido, Japan</td>
<td>A</td>
<td>AB361611*</td>
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<tr>
<td>A. ramosa Franch.</td>
<td>RAM9</td>
<td>Mt. Fujiwara, Mic, Japan</td>
<td>B(1ID)</td>
<td>AB361618*</td>
</tr>
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<td>A. ramosa Franch.</td>
<td>RAM10</td>
<td>Ina, Nagano, Japan</td>
<td>B(1ID)</td>
<td>AB361616*</td>
</tr>
<tr>
<td>A. ramosa Franch.</td>
<td>RAM11</td>
<td>Yonezawa, Yamagata, Japan</td>
<td>A</td>
<td>AB361614*</td>
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<tr>
<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE1</td>
<td>Saengil Island, Jeonnam, Korea s.n. (HNHM)</td>
<td>M(16NS)</td>
<td>KU570398</td>
</tr>
<tr>
<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE2</td>
<td>Donghae-si, Gangwon, Korea s.n. (HNHM)</td>
<td>N(16NS)</td>
<td>KU570399</td>
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<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE3</td>
<td>Pung Island, Gyeonggi, Korea s.n. (HNHM)</td>
<td>O(16NS)</td>
<td>KU570400</td>
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<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE4</td>
<td>Hyoganchi, Jeonbuk, Korea s.n. (HNHM)</td>
<td>G(14NS)</td>
<td>KU570401</td>
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<td>A. pseudoamurensis W. T. Wang</td>
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<td>Jangbong Island, Gyeonggi, Korea</td>
<td>H(14NS)</td>
<td>AF454935*</td>
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<tr>
<td>A. pseudoamurensis W. T. Wang</td>
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<td>Euyungi, Gyeongnam, Korea</td>
<td>J(15NS)</td>
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<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE7</td>
<td>Mt. Palgong, Gyeongbuk, Korea</td>
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<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE8</td>
<td>Gwanchon, Jeonbuk, Korea</td>
<td>G(14NS)</td>
<td>AF454931*</td>
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<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE9</td>
<td>Mt. Gyeryong, Chungnam, Korea</td>
<td>K(15NS)</td>
<td>AF454933*</td>
</tr>
<tr>
<td>A. pseudoamurensis W. T. Wang</td>
<td>PSE10</td>
<td>Chollipo, Chunchnam, Korea</td>
<td>K(15NS)</td>
<td>AF454934*</td>
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<tr>
<td>A. multiflora Nishikawa et Koji Ito</td>
<td>MUL1</td>
<td>Mulchatorum, Jeju, Korea s.n. (HNHM)</td>
<td>L(15NS)</td>
<td>KU570395</td>
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<tr>
<td>A. multiflora Nishikawa et Koji Ito</td>
<td>MUL2</td>
<td>Seondeoksa, Jeju, Korea s.n. (HNHM)</td>
<td>L(15NS)</td>
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<td>A. multiflora Nishikawa et Koji Ito</td>
<td>MUL4</td>
<td>Sangumburi, Jeju, Korea</td>
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<td>AF454926*</td>
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<tr>
<td>A. multiflora Nishikawa et Koji Ito</td>
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<td>Eorimok, Jeju, Korea</td>
<td>G(14NS)</td>
<td>AF454925*</td>
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<td>MUL6</td>
<td>Kawai, Gifu, Japan</td>
<td>I(15NS, 1ID)</td>
<td>AB361621*</td>
</tr>
</tbody>
</table>
Phylogenetic study of the section Adonanthe of genus Adonis L. (Ranunculaceae) based on ITS sequences

Crandall, 1998). After the best model was selected, $2 \times 10^6$ generations were run with the Markov chain Monte Carlo method (MCMC; Hastings, 1970) using MrBayes ver. 3.1.2 (Huelsenbeck and Ronquist, 2001), and then posterior probabilities were calculated after discarding the burn-in sample (initial 500,000 generations).

Furthermore, a tree topology built using the Neighbor Joining method (NJ) (Saitou and Nei, 1987; Farris et al., 1995) and pairwise sequence divergence was calculated by the Kimura two-parameter method (Kimura, 1980).

Results

Sequences of nrDNA ITS region

The total DNA sequence length of the ITS region from 49 accessions representing 13 taxa in the sect. Adonanthe was 597–601 bp, and the total length of the aligned sequences was 604 bp. The length of ITS 1 and ITS 2 was 232–234 bp and 201–203 bp, respectively, showing that the length of ITS 1 was longer than that of ITS 2. The length of the 5.8S region was 164–165 bp, which was similar to that in other flowering plants (161–164 bp).

Pairwise distance between taxa calculated using Kimura's two parameter method revealed that sequence divergence of the ITS regions across 53 samples, including the outgroup, was 0–9.873%; and sequence divergence among 49 samples, excluding the outgroup, was relatively high (0–6.106%). In the data set that included outgroups, taxa without intraspecific sequence variation included A. ramosa (RAM1, RAM3, RAM4, RAM5, RAM7, RAM8, and RAM11), A. amurensis (AMU6 and AMU7), A. shikokuensis (SHI1), A. pseudoamurensis (PSE4, PSE7, and PSE8), and A. multiflora (MUL4 and MUL5) (Table 1). The highest sequence divergence was observed between A. annua and A. amurensis (AMU1). Among the ingroups, A. davidii (DAV1) and A. amurensis (AMU3 and AMU5) were the taxa with the highest sequence divergence (6.106%).

Phylogenetic analyses

NJ analysis based on sequence divergence resolved the species of the Adonis sect. Adonanthe largely into four clades (Fig. 1). Clade I comprised A. sutchuenensis (Franch.), A. davidii, and A. brevistylo from ser. Amurense and A. coerulea and A. coerulae var. puberula from ser. Coeruleae; it was resolved as the most basal, forming a sister group with other clades. Then A. vernalis from ser. Vernales (clade II) diverged, and clade III including A. bobroviana Simonovich and A. sibirica (Patrin ex DC.) Patrin ex Ledeb. from ser. Apenninae diverged after A. vernalis. Clade IV comprised two subclades, subclade 1 consisting of A. pseudoamurenensis, A. multiflora, and A. shikokuensis, and subclade 2 including A. amurenensis, and A. ramosa, which is congruent with the phylogenetic tree inferred from MP analysis (Fig. 2). Within the subclade 1, some accessions (PSE4, PSE7, PSE8, MUL4, and MUL5) possessed the same ITS sequences with A.
Also, two accessions (AMU6 and AMU7) of *A. amurensis* were located in the same subclade as *A. ramosa*, showing a genetically close relationship between these taxa.

MP analysis of the ITS regions resulted in 34 equally parsimonious trees with a tree length of 170 steps, in which the CI (Consistency Index), RI (Retention Index), and RC (Rescaled Consistency Index) were 0.800, 0.931, and 0.744, respectively. The total length of the aligned sequences was 604 bp, of which, 115 were variable sites and 76 were parsimony informative sites.

In the parsimony tree (Fig. 2), the sect. *Adonanthe* of genus *Adonis* L. formed a monophyletic group consisting of 4 clades. The sect. *Adonanthe* had bootstrap values less than 50% for divergence of a clade (clade I) composed of *A. sutchuenensis*, *A. coerulea*, *A. coerulea* var. *puberula*, *A. davidii*, and *A. shikokuensis* (Table 1). Also, two accessions (AMU6 and AMU7) of *A. amurensis* were located in the same subclade as *A. ramosa*, showing a genetically close relationship between these taxa.

Fig. 1. Neighbor-Joining (NJ) tree inferred from nrDNA ITS sequences of the section *Adonanthe* under genus *Adonis* and its related genera by Kimura-two parameter distance. The numbers on branches indicate bootstrap values higher than 50%. Refer to Table 1 for accession abbreviation.
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In this clade, *A. sutchuenensis* formed a subclade with an 88% bootstrap value, and a sister subclade composed of *A. davidii*, *A. brevistyla*, *A. coerulae*, and *A. coerulae* var. *puberula* was formed. *A. vernalis* (clade II) diverged with a 100% bootstrap value, and a clade III composed of *A. bobroviana* and *A. sibirica* diverged with a bootstrap value less than 50%. In addition, a subclade composed of *A. pseudoamurensis*, *A. multiflora*, and *A. shikokuensis* and a subclade composed of *A. amurensis* and *A. ramosa* formed clade IV with a bootstrap value of 65%. Ser. *Amurenses* formed a polyphyletic group with representatives present in clade I and clade IV.

ML analysis (Fig. 3) showed monophyly of the genus *Adonis* sect. *Adonanthe* with bootstrap values of 100% and a Bayesian posterior probabilities value of 1.00. The clade I observed on the NJ and MP tree was not evident in the ML tree. *A. vernalis* diverged with a bootstrap value of 100% and a Bayesian posterior probabilities value of 1.00 (clade II). Clade III
composed of *A. bobroviana* and *A. sibirica* formed a sister group with clade IV composed of *A. pseudoamurensis*, *A. multiflora*, *A. shikokuensis*, *A. amurensis*, and *A. ramosa* with a bootstrap value of 92% and a Bayesian posterior probability value of 0.99. A subclade composed of *A. amurensis* and *A. ramosa* within clade IV formed a sister group with a subclade composed of *A. pseudoamurensis*, *A. multiflora*, and *A. shikokuensis* with a bootstrap value of 63% and a Bayesian posterior probability value of 0.99.

**Discussion**

A phylogenetic study using ITS sequences was conducted for 13 taxa that belong to the sect. *Adonanthe* of genus *Adonis* provided valuable information for the classification of the taxonomically problematic section. Although the ITS trees mostly supported monophyly of ser. *Vernales* and ser. *Apenninae*, the taxa in ser. *Amurenses* did not form a monophyletic group (Figs. 1–3). The polyphyly of ser.
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Adonanthe is probably due to genetic divergence caused by disjunctive distribution of the members in the series (A. davidii, A. brevistyla, and A. sutchuenensis in mid-western China and other taxa in northeastern China, Korea, and Japan). These results are inconsistent with the classification system proposed by Wang (1994a, 1994b).

Meanwhile, the taxa in the clade I (Figs. 1, 2), which is composed of A. davidii, A. brevistyla, and A. sutchuenensis in ser. Amurenses and A. coerulea and A. coerulea var. puberula in ser. Coeruleae, grow in fruticeta or grasslands at an elevation of 1,500–3,000 m in the mid-southern, southwestern, and western areas in China and are characterized by elongated and lignified rhizomes. This is in contrast with those taxa found in northeastern China, Korea, and Japan, including the distinct clade IV (A. amurensis, A. ramosa, A. pseudamurensis, A. multiflora, and A. shikokaiensis), which have short rhizomes and are surrounded by fibrous roots. In the strict consensus tree (Fig. 2), however, clade I composed of A. sutchuenensis, A. coerulea, A. coerulea var. puberula, A. davidii, and A. brevistyla collapsed, which is consistent with the results from the ML analysis (Fig. 3). Despite forming a clade in the NJ analysis (Fig. 1), either the bootstrap value supporting the clade I was low or the taxa examined did not form a monophyletic group depending on the analysis method implemented. Thus, further studies are needed to resolve the phylogenetic relationships among these groups in relation to the evolutionary trends of morphological characters.

There have been many different opinions regarding the taxonomic identities and phylogenetic relationships of the taxa that belong to the sect. Adonanthe. It is known that the members of sect. Adonanthe have similar morphological characters with severe variations, which hindered establishment of dependable taxonomic characters within the section. A. davidii and A. brevistyla formed a clade with bootstrap value of 99% in the MP analysis (Fig. 2), which revealed that they are genetically related taxa. It is noticeable that BRE1 and BRE2 in A. brevistyla did not form a clade in all ITS trees (Figs. 1–3). Depending on the taxonomists, A. davidii and A. brevistyla have been considered either conspecific (Wang, 1980; Fu, 2000; Fu and Robinson, 2001) or treated as independent species (Wang, 1994a). Since it is possible that A. davidii was misidentified as BRE1 in the present study, a taxonomic study based on the original description and a type specimen should be performed to validate the identities of these taxa. Meanwhile, Wang (1994a, 1994b) suggested that A. davidii was the most primitive species in the sect. Adonanthe. However, clade I collapsed in the strict consensus tree, making the group consisting of A. davidii, A. sutchuenensis, A. coerulea, and A. coerulea var. puberula is paraphyletic, which is inconsistent with the taxonomic opinion of Wang (1994a, 1994b). Nevertheless, A. davidii is positioned at the base of sect. Adonanthe in MP, NJ, and ML analyses (Figs. 1–3). In addition, the species has ancestral characters, including long rhizomes, long petioles, oval lobules, and short persistent styles with recurved ends and is found in the southwestern region of China. This region is the center of origin for the genus Adonis L., and for this reason it has been considered to be the progenitor species of the sect. Adonanthe (Wang, 1994a, 1994b; Son, 2015).

Adonis coerulea and A. coerulea var. puberula were clustered together in the NJ analysis (Fig. 1), yet they did not form a clade in MP and ML trees (Figs. 2, 3). In some cases, A. coerulea var. puberula has been considered as a synonym of A. coerulea by Fu and Robinson (2001); thus, use of multiple samples collected from diverse areas and employment of phylogenetically useful DNA regions is imperative in order to confirm their taxonomic identities.

Although A. sutchuenensis, an endemic Chinese species, is superficially similar to A. amurensis, it is clearly distinguished by following morphological characters: the leaf shape is ovate pentagon and the sepal is shorter and narrower than the petal (Franchet, 1894). Within the ITS trees, multiple accessions representing A. sutchuenensis formed a strong monophyletic group, supporting the species is clearly distinct from other taxa within the sect. Adonanthe; therefore, it is reasonable to treat it as an independent species.

Adonis vernalis formed a sister clade with the group composed of clade IV (including representatives in ser. Amurenses found in northeastern China, Korea, and Japan) and clade III (ser. Apenninae distributed in mid-western China, Mongolia, Russia, and Kazakhstan), indicating that it is directly related to these groups (Figs. 1–3). In the sect. Adonanthe, A. vernalis is the only taxon distributed in Europe and Central Asia; therefore, it is thought that A. vernalis diverged and formed a distinct species when it moved from the western Himalayas to Europe. This divergence within the genus Adonis L. appears to support the views that the plants of the sect. Adonanthe (excluding A. vernalis) underwent speciation either after being transmitted along the route from southwestern China to Eastern Siberia via the northeastern Chinese region or from southwestern China to North Asia (Wang, 1994a, 1994b; Son, 2015).

Although the support for the clade is not sufficient, all ITS trees recognized monophyly of ser. Apenninae, which is supported by distinctive morphological characters such as lack of petioles (all other taxa of the sect. Adonanthe have petioles). Phylogenetic analysis, however, revealed that A. bobroviana is not monophyletic. The morphological characteristics of A.
Adonis ramosa was treated either as a Japanese endemic species (Nishikawa and Kadota, 2006) or the same species as *A. pseudoamurensis*, which is found in Korea and China (Wang and Liu, 1988; Wang, 1994a, 1998; Fu and Robinson, 2001; Park, 2007). Additionally, *A. pseudoamurensis* was considered to be the same species as *A. multiflora* found in Korea and Japan (also reported in China; Nishikawa and Kadota, 2006); thus, there has been confusion concerning taxonomic identities of the taxa. The MP, NJ, and ML analyses of the ITS sequences distinguished *A. ramosa* from *A. pseudoamurensis* and revealed that *A. multiflora* was a distinct clade (Figs. 1–3). Morphologically, *A. ramosa* has hairs on the abaxial side of the leaf and its sepal length is similar to that of the petals. In contrast, *A. pseudoamurensis* and *A. multiflora* have no hair on the abaxial side of the leaves and the sepals are shorter than the petals. In addition, since *A. pseudoamurensis* has distinctive traits from *A. multiflora* with respect to the morphology of the leaf apex and lobule and microstructure of the achene surface (Son and Ko, 2012, 2013), it is reasonable to treat these species as independent taxa. With respect to chromosome number in the somatic cells, *A. ramosa* is a tetraploid plant with 2n = 32 (Nishikawa, 1988, 1989), whereas *A. pseudoamurensis* is a diploid plant with 2n = 16 (Son, 2011). Thus, it is unlikely that *A. ramosa* is distributed in Korea and China (Wang and Liu, 1988; Wang, 1994a, 1998; Fu and Robinson, 2001; Park, 2007). Instead, it appears to be reasonable to consider the species to be a Japanese endemic species (Suh et al., 2002; Lee et al., 2003; Nishikawa and Kadota, 2006). On the other hand, *A. ramosa* and *A. amurensis* formed a subclade with a bootstrap value of 75% in the MP tree (Fig. 2), indicating they are closely related taxa. However, the Korean *A. amurensis* and Japanese *A. amurensis* groups formed distinctive subclades. Interestingly, the Japanese *A. amurensis* accessions and the accessions of *A. ramosa* (RAM1, RAM3, RAM4, RAM5, RAM7, RAM8, and RAM11) had identical DNA sequences (Table 1). Geographically, *A. ramosa* is distributed in Hokkaido and northern to mid Honshu (Nishikawa and Kadota, 2006), and this area overlaps with that of *A. amurensis* in the eastern Hokkaido region (Nishikawa and Takeshima, 1992; Nishikawa, 1994). This is consistent with the theory of Kaneko et al. (2008), which speculated that *A. ramosa* should be an autotetraploid and the diploid progenitor of *A. ramosa* should be *A. amurensis* from the Hokkaido region. Since only 2 ITS ribotypes in *A. ramosa* were examined (Table 1), the data may not be sufficient to determine the process of speciation. However, the geographical distribution of organisms related to their ITS ribotypes was clearly distinct (Kaneko et al., 2008), implying the identical DNA sequence between the Japanese *A. amurensis* and *A. ramosa* appears to be due to a relatively recent speciation event.

For the Korean *A. amurensis*, the MP, NJ, and ML analyses divided the group into two subgroups (Figs. 1–3), which was attributed to the differences in the insertion/deletion of sequences (Table 1). However, no clear relationship was observed between the geographical distributions of these individuals and the ribotypes of each sample.

The ITS sequence analysis of *A. pseudoamurensis* revealed high levels of variation within the species (Table 1). This result is consistent with the report of Suh et al. (2002) that *A. pseudoamurensis*, unlike *A. amurensis* and *A. multiflora*, showed a high level of genetic variation in RAPD analysis. The high genetic variation in *A. pseudoamurensis* is probably the result of adaptation to diverse habitats in wide geographical areas of the species which ranges from northeastern China to southern part of the Korean Peninsula including islands on the west to south coast as well as lowland regions. Meanwhile, ITS trees indicated that *A. pseudoamurensis* is clearly distinct from *A. amurensis* (Figs. 1–3). In morphology, *A. amurensis* has an unbranched stem and its leaves emerge after the inflorescence has developed, whereas *A. pseudoamurensis* has a branched stem and its leaves and flowers develop at the same time; these differences together with phylogenetic data further validate recognition of *A. pseudoamurensis* as an independent species (Lee et al., 2003; Son and Ko, 2011, 2012) rather than as a variety of *A. amurensis* (Lee, 2004, 2006).

Some accessions of *A. multiflora* formed a subclade with a bootstrap value of 63% in MP analysis (Fig. 2). Additionally, the Sangumburi (MUL4) and Eorimok (MUL5) accessions were similar to *A. shikokuensis* that is distributed in Honshu, Shikoku, and Kyushu in Japan as well as to other groups (PSE4, PSE7, and PSE8) of the Korean *A. pseudoamurensis* (Figs. 2, 3, Table 1). This result suggest that *A. shikokuensis* might be related with either *A. pseudoamurensis* or *A. multiflora*, contradicting the hypothesis proposed by Nishikawa and Ito (2001), who suggested *A. shikokuensis* diverged from *A. ramosa*. However, further studies including more diverse samples and additional molecular markers are necessary to resolve the relationship among these taxa and obtain a reliable phylogenetic tree.
Acknowledgments

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Literature Cited


Son, D. C. 2011. A taxonomic study on the genus Adonis L. (Ranunculaceae) in Korea. Hannam University, Daejeon. (in Korean)

Son, D. C. 2015. A systematic study on the section Adonanthae of genus Adonis L. (Ranunculaceae) in East Asia. Hannam University, Daejeon. (in thesis for D. Sc.)


Son, D. C. 2015. A systematic study on the section Adonanthe of genus Adonis L. (Ranunculaceae) in East Asia. Hannam University, Daejeon. (in thesis for D. Sc.)

