Taxonomic reconsideration of Chinese *Lespedeza maximowiczii* (Fabaceae) based on morphological and genetic features, and recommendation as the independent species *L. pseudomaximowiczii*

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**ABSTRACT:** *Lespedeza maximowiczii* C. K. Schneid. (Fabaceae) is a deciduous shrub which is known to be distributed in the temperate forests of China, Korea and on Tsushima Island of Japan. Due to severe morphological variations within species, numerous examinations have been conducted for Korean *L. maximowiczii*. However, the morphology of Chinese plants has not been studied as thoroughly, despite doubts about their taxonomy. To clarify this taxonomic issue, we investigated morphological characters and undertook a Bayesian clustering analysis with microsatellite markers. The morphological and genetic traits of Chinese individuals varied considerably from those of typical *L. maximowiczii* growing in Korea. For example, petals of the former had a different shape and bore long claws, while the calyx lobes were diverged above the middle and the upper surface of the leaflet was pubescent. Their terete buds and spirally arranged bud scales were distinct from those within the series/section *Heterolespedeza*, which includes *L. maximowiczii*. Our Bayesian clustering analysis additionally included *L. buergeri* as an out-group. Those results indicated that the Chinese samples clustered into a lineage separated from *L. maximowiczii* (optimum cluster, *K* = 2), despite the fact that the latter is grouped into the same lineage with *L. buergeri*. Therefore, we treat those Chinese plants as a new species with the name *L. pseudomaximowiczii*.

**Keywords:** Bayesian clustering analysis, *Lespedeza maximowiczii*, *Lespedeza pseudomaximowiczii*, microsatellite, morphology, new species

*Lespedeza maximowiczii* C. K. Schneid. is a deciduous shrub in the family Fabaceae (Choi, 2007). This species is traditionally considered to belong to section (Nakai, 1939) or series (Akiyama, 1988) *Heterolespedeza* Nakai, which is characterized by flattened buds and distichously arranged bud scales (Nakai, 1939; Akiyama, 1988). In a recent revision of the taxonomic system for *Lespedeza* Michx., (Ohashi and Nemoto, 2014), *Heterolespedeza* was placed in section *Macrolespedeza* Maxim., reflecting the results of molecular phylogenetic studies (Han et al., 2010; Xu et al., 2012). Simultaneously, series *Formosae* S. Akiyama & H. Ohba (including *L. thunbergii* (DC.) Nakai, *L. davidii* Franch., *L. homoloba* Nakai, and *L. patens* Nakai) was treated as a synonym of section *Macrolespedeza* (Ohashi and Nemoto, 2014). First described based on Faurie’s collection (Schneider, 1907), *L. maximowiczii* was sampled from Quen-san (= Wonsan-si), Hamgyeongnam-do, Korea. The origins of its local name were reviewed by Chang et al. (2004). Now, this species is known to be distributed in the temperate forests of Korea, China, and Japan (Akiyama, 1988; Choi, 2007; Ohashi et al., 2009). Plants grow abundantly on the mountains in all Korean provinces (Choi, 2007), and are also found in Anhui, Henan and Zhejiang provinces of central China (Ohashi et al., 2009). In Japan, this species is restricted to the northern part of Isl. Tsushima (Akiyama, 1988), close to the Korean Peninsula. Therefore, the highest concentrations of plants are in China

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and Korea, two regions separated by ca. 1170 km.

The morphological characters of *L. maximowiczii* vary widely, with many intra taxa being described, such as var. *tomentella*, var. *elongata*, and f. *alba* (Nakai, 1927; Akiyama, 1988). To clarify the circumscription on this species, those characters have been examined several times for Korean and Japanese individuals (Lee, 1965; Hatusima, 1967; Akiyama, 1988), and the taxonomic rank of this species and intra taxa has also been changed by researchers. The Japanese individuals are regarded as typical forms even though some have smaller calyces (ca. 2 mm long) (Hatusima, 1967; Akiyama, 1988). However, only a few morphological investigations have been performed with Chinese *L. maximowiczii*. Although Ohashi et al. (2009) and Huang et al. (2010) revised Chinese *Macrolespedeza*, shrub forms of *Lespedeza*, they did not consider *L. maximowiczii* when observing those specimens. Flower characters are key to distinguishing among *Lespedeza* species (Akiyama, 1988) but have not been described in minute detail. If plants of that species have been geographically isolated between China and Korea/Japan for a long time, it is possible that Chinese individuals show distinct morphological traits and should be treated as intra taxa or a new species. In the case of the related *L. buergeri* Miq., which belongs to the same series/section *Heterolespedeza*, plants in the Korean population differ from those of Chinese and Japanese populations because of their smaller bracteoles (Jin et al., 2016b). Hence, morphological examination of Chinese individuals is required to confirm its taxonomic entity.

Several phylogenetic studies on *Lespedeza* have utilized molecular markers such as cpDNA and nrITS (Han et al., 2010; Xu et al., 2012), and nrITS and nuclear gene PGK (Xu et al., 2017). However, Chinese and Korean *L. maximowiczii* have never been sequenced in parallel. It is difficult to delimit species of *Macrolespedeza* because the resolution of markers is low due to frequent hybridization, reticulate evolution, and rapid diversification (Han et al., 2010; Xu et al., 2012, 2017). The PGK markers show relatively high resolution for section *Junceae* (Maxim.) H. Ohashi & T. Nemoto but not for *Macrolespedeza*. Therefore, other markers are needed. We employed microsatellite markers developed from this genus (Jin et al., 2016a). Because they are highly polymorphic and distributed across multiple loci (Duminil et al., 2012), they can be used to reveal genetic diversity among populations (Sunnucks, 2000) as well as to delimit closely related taxa in various groups, such as those within the genera of *Carapa* Aubl., *Phoenix* L. and *Quercus* L. (Pintaud et al., 2010; Duminil et al., 2012; Lee et al., 2014).

Here, our research aims were to (1) compare the morphological and genetic traits of typical *L. maximowiczii* with those of Chinese samples currently identified as *L. maximowiczii* and (2) clarify the taxonomic entity of plants growing in China.

**Materials and Methods**

**Morphological examination and survey of geographical distribution**

To compare the morphological characters of typical *Lespedeza maximowiczii* with those of Chinese plants identified the same way, we collected specimens from both countries and deposited them in the Herbarium of Inha University (IUI). Additional morphological examination was conducted by observing specimen from the following herbaria: the Korea National Arboretum (KIH) and the Chinese Academy of Sciences, Beijing (PE). We also gathered more information throughout the Chinese Virtual Herbarium (CVH) (http://www.cvh.ac.cn/); and by looking at photograph and geographical distribution of Chinese specimens held at the Hangzhou Botanical Garden (HHBG); the Institute of Botany, Jiangsu Province and Chinese Academy of Sciences (NAS); and the Kunming Institution of Botany (KUN). All observations of morphological characters were made with a stereomicroscope (Leica MZ8; Wetzlar, Germany), and criteria for floral measurements were mainly those stipulated by Akiyama (1988).

**Sampling, DNA extraction, and microsatellite polymerase chain reaction**

Genetic traits were studied using leaves sampled from typical forms (Korean, *n* = 87) and Chinese individuals (*n* = 38) which were identified as *L. maximowiczii* (Table 1). Plants of *L. maximowiczii* also grow, albeit only in small numbers, on Isl. Tsushima (Akiyama, 1988). Because of that scarcity, we did not include them in this molecular examination. As the outgroup, we analyzed one population each from China and Japan of the related species *L. buergeri* (*n* = 14 and *n* = 11, respectively).

Genomic DNA (3 μg) was extracted from silica-dried leaves with an MG Plant Genomic DNA Extraction SV Miniprep Kit (MGmed, Seoul, Korea), according to the manufacturer’s instructions. For Bayesian cluster analysis, we used the following eight microsatellite loci (Jin et al., 2016a): LMS3, LMS18, LMS28, LMS39, LMS45, LMS47, LMS53, and LMS58. The polymerase chain reaction (PCR) protocol utilized those markers with a GeneAmp PCR System 2700 Thermal Cycler (Applied Biosystems, Foster City, CA, USA). Each reaction mixture (10 μL total volume) contained 5 ng of DNA,
plus 5 μL of 2× Plus Mix (Dongsheng Biotech, Guangdong, China) that comprised 0.4 mM dNTPs, 2× PCR buffer with 4 mM MgSO$_4$, and 0.4 U/μL of Taq DNA polymerase. The mixtures also contained the appropriate 0.08 μM forward M13 (-21)-tagged primer, a 0.3 μM reverse primer, and a 0.3 μM M13 (-21) labeled fluorescent marker (NED, PET, VIC, 6-FAM). Conditions included initial denaturation at 94°C for 3 min; then 30 cycles at 94°C for 30 s, 53°C for 30 s, and 72°C for 45 s; without a final extension. Afterward, the PCR products were visualized on 2% agarose gels and resolved to genotype on an ABI 3730XL sequencer with GeneScan 500 LIZ size standards (Applied Biosystems). The sizes of the alleles were determined with GENEMAPPER 3.7 (Applied Biosystems).

**Microsatellite data analyses**

Based on genotype data, we analyzed the genetic characters in terms of the number of alleles ($N_a$), number of private alleles ($N_p$), and expected heterozygosity ($H_e$) with GenAlEx 6.5.3 (Peakall and Smouse, 2012). Because the Bayesian clustering method assigns ancestral lineages for all individuals, we were able to use it to compare the genetic structures of Korean and Chinese samples. This analysis was performed with STRUCTURE 2.3.4 (Pritchard et al., 2000), selecting the admixture ancestry and correlated allele frequency models. Ten runs were conducted for each cluster ($K = 1$ to 16), with 200,000 burn-ins followed by 200,000 Markov chain Monte Carlo repetitions. To determine the optimum $K$ values, we calculated $\Delta K$ using STRUCTURE HARVESTER (Earl and von Holdt, 2012).

**Results and Discussion**

**Morphological examination of Chinese plants identified as Lespedeza maximowiczii**

To confirm the taxonomic entity of Chinese plants currently identified as *Lespedeza maximowiczii*, we compared their morphological traits with those of Korean *L. maximowiczii* (Fig. 1, Table 2). Our main examination focused on the external morphology of buds, flowers, and leaflets. To avoid any misidentification of individuals collected for this study, we tried to observe the specimen (collection no. K.M. Liu 4846) that had been used originally to report the occurrence of *L. maximowiczii* in China (Kung, 1936). However, despite those attempts, we could not observe any such specimen from the CVH. Therefore, we looked at specimens from the same date and site collected by the same person (collection no. K.M. Liu 4864) in the PE herbarium. This enabled us to determine the morphological traits of Chinese plants while preventing any erroneous identifications.

The differences between Chinese plants and typical *L. maximowiczii* are summarized in Table 2. Although flower lengths were similar, i.e., 9.1–(9.8)–10.5 mm for Chinese individuals versus 8.2–(9.5)–10.9 mm for the Korean samples, the ratio of petal claw to lamina length varied. In particular, the standard petals of Chinese plants showed relatively longer claws (2.4–[2.7]–3.0 mm) than those of the typical specimen (0.7–[1.1]–1.5 mm). The standards were narrower for Chinese samples (4.1–[4.8]–5.5 mm vs. 4.4–
Fig. 1. Comparison of morphological traits between typical *Lespedeza maximowiczii* and Chinese plant, named *L. pseudomaximowiczii*. A. Korean *L. maximowiczii*. B. *L. pseudomaximowiczii*. a, bud; b, hairs on leaflet; c, standard petal; d, wing petal; e, keel petal; f, calyx; g, flower.
Table 2. Comparison of morphological traits between typical Lespedeza maximowiczii and Chinese plant identified as L. maximowiczii.

<table>
<thead>
<tr>
<th>Trait</th>
<th>Typical L. maximowiczii</th>
<th>Chinese plant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Shape of bud</td>
<td>Mostly flattened</td>
<td>Terete</td>
</tr>
<tr>
<td>Arrangement of bud scales</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Length of flower (mm)</td>
<td>8.2 (9.5)–10.9</td>
<td>9.1 (9.8)–10.5</td>
</tr>
<tr>
<td>Length of standard petal (mm)</td>
<td>7.0 (8.5)–10.0</td>
<td>8.7 (9.3)–10.5</td>
</tr>
<tr>
<td>Length of standard petal claw (mm)</td>
<td>0.7 (1.1)–1.5</td>
<td>2.4 (2.7)–3.0</td>
</tr>
<tr>
<td>Width of standard petal claw (mm)</td>
<td>4.4 (5.6)–6.8</td>
<td>4.1 (4.8)–5.5</td>
</tr>
<tr>
<td>Length of keel petal (mm)</td>
<td>5.6 (8.5)–12.0</td>
<td>3.1 (3.5)–4.0</td>
</tr>
<tr>
<td>Length of wing petal lamina (mm)</td>
<td>5.6 (6.3)–7.0</td>
<td>4.3 (5.4)–6.2</td>
</tr>
<tr>
<td>Length of wing petal lamina (mm)</td>
<td>6.2 (6.3)–7.0</td>
<td>4.3 (5.4)–6.2</td>
</tr>
<tr>
<td>Length of keel petal lamina (mm)</td>
<td>6.3 (6.3)–7.0</td>
<td>4.3 (5.4)–6.2</td>
</tr>
<tr>
<td>Length of keel petal lamina (mm)</td>
<td>7.0 (7.0)–10.0</td>
<td>3.3 (3.8)–4.4</td>
</tr>
<tr>
<td>Length of standard petal claw (mm)</td>
<td>1.7 (2.4)–2.9</td>
<td>3.1 (3.5)–4.0</td>
</tr>
<tr>
<td>Length of wing petal lamina (mm)</td>
<td>2.4 (2.4)–3.3</td>
<td>2.2 (2.4)–2.8</td>
</tr>
<tr>
<td>Length of keel petal lamina (mm)</td>
<td>2.2 (2.2)–3.2</td>
<td>2.2 (2.4)–2.8</td>
</tr>
<tr>
<td>Length of standard petal claw (mm)</td>
<td>1.5 (2.3)–3.5</td>
<td>1.9 (2.0)–2.2</td>
</tr>
</tbody>
</table>
| Density of hairs on upper surface of leaflet (number per 4 mm²) | 1 (8)–59 | 86 (144)–218

Number in parenthesis indicates mean value.

[5.62]–6.8 mm and they also varied in shape. Whereas those of the Chinese plants were oblong or elliptic, the typical form was elliptic to broad elliptic. For Korean samples, the ratio of claw to lamina length in the wing and keel petals (wing, 1:1.6–[2.3]–2.8; keel, 1:1.7–[2.5]–3.3) closely coincided with previous observations (ratio of ca. 1:2.5 in both petal types) (Akiyama, 1988). This contrasted with length ratios for the Chinese wing (1:1.2–[1.3]–1.6) and keel (1:1.0–[1.4]–1.6). The calyx from Chinese individuals was lobed above the middle portion (tube, 2.2–[2.4]–2.8 mm long; lobe, 1.9–[2.0]–2.2 mm long), whereas the calyx of the typical form was usually lobed below the middle portion (tube, 1.2–[1.5]–2.1 mm long; lobe, 1.5–[2.3]–3.5 mm long). The density of hairs on the upper surface of a leaflet (defined as the number of hairs per 4 mm²) was much higher for the more pubescent Chinese plants (ca. 144) than for the typical L. maximowiczii (ca. 8). Previous examinations have demonstrated that even the leaflets of Korean L. maximowiczii f. tomentella are pubescent up until the fruiting season (Nakai, 1927, Akiyama, 1988). We observed their density of hairs on the upper surface of a leaflet, and it is lower than that of Chinese plants (ca. 51). We also could not regard the Chinese individuals as part of Heterolespedeza because they had terete buds and spirally ordered bud scales (a in Fig. 1B). Therefore, these traits of buds and flowers, considered an important key for identifying Lespedeza species (Akiyama, 1988; Ohashi et al., 2009), were sufficient to suggest that the Chinese individuals are a distinct species.

**Genetic characters and Bayesian-clustering based on eight microsatellite loci**

We genotyped 150 individuals of typical L. maximowiczii (n = 87), Chinese plants (n = 38), and L. buergeri (n = 25) based on our microsatellite loci. Most of the eight loci were highly variable for all taxa (Table 3). In particular, LMS18 (He = 0.753–0.825) and LMS39 (He = 0.642–0.879) showed high genetic diversity while LMS53 (He = 0.000–0.301) harbored low diversity. The degree of diversity also differed according to taxa, with LMS28 showing a high value in the Chinese samples (He = 0.694) but producing low values in the typical L. maximowiczii (He = 0.203) and L. buergeri (He = 0.000). For LMS45, the Chinese plants presented a lower value (He = 0.193) than that calculated for the Korean L. maximowiczii (He = 0.595) and L. buergeri (He = 0.619). Our analysis of genetic structure among species (Fig. 2) indicated an optimum cluster of two because ΔK was highest at K = 2. Bayesian clustering results (K = 2) showed that typical L. maximowiczii was grouped in the same cluster with L. buergeri while the Chinese individuals were assigned to an independent lineage (Fig. 3). This might have resulted because of geographic isolation rather than delimitation among species. However, it was not unexpected because Chinese plants were distinct from typical L. maximowiczii, which were, in turn, closer to L. buergeri than to the Chinese plants. At K = 3 that showed second highest ΔK value, L. maximowiczii and L. buergeri were distinguished as separate species rather than being populations within the same species. In both K = 2 and 3, most of individuals of
typical *L. maximowiczii*, Chinese plant, and *L. buergeri* are well assigned to one major cluster, but some individuals exhibited a mixed lineage (posterior probabilities that originates from major cluster \( q \) < 0.9). This mixture is perhaps due to a genetic exchange among *Lespedeza* species, as has been suggested previously (Xu et al., 2012, 2017). In fact, this assumption is supported by reports of many hybrids in *Lespedeza*, based on morphological examinations (e.g., Lee, 1965; Akiyama and Ohba, 1982, 1983; Akiyama, 2004). Although those individuals might have resulted from hybridization or introgression, genetic variations within the population could also be a factor. Because of those vagaries, we did not determine their origin(s) in our current exploration.

**Taxonomic entity for Chinese plant**

Chinese plant was originally identified as *Lespedeza maximowiczii* due to morphological similarities. However, our results demonstrated that those individuals should be classified as a distinct species. To confirm this, we compared their morphological traits with other species that grow in China. Because of their terete buds and spiral arrangement of bud scales (Nakai, 1939; Ohashi et al, 2009), it can be difficult from major cluster \( q \) < 0.9). This mixture is perhaps due to a genetic exchange among *Lespedeza* species, as has been suggested previously (Xu et al., 2012, 2017). In fact, this assumption is supported by reports of many hybrids in *Lespedeza*, based on morphological examinations (e.g., Lee, 1965; Akiyama and Ohba, 1982, 1983; Akiyama, 2004). Although those individuals might have resulted from hybridization or introgression, genetic variations within the population could also be a factor. Because of those vagaries, we did not determine their origin(s) in our current exploration.

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**Table 3.** Genetic characteristics of *Lespedeza maximowiczii* (China vs. typical) and *L. buergeri* based on 8 microsatellite loci.

<table>
<thead>
<tr>
<th>Locus</th>
<th>Na</th>
<th>Np</th>
<th>He</th>
<th>Range</th>
<th>Na</th>
<th>Np</th>
<th>He</th>
<th>Range</th>
<th>Na</th>
<th>Np</th>
<th>He</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>LMS3</td>
<td>8</td>
<td>2</td>
<td>0.708</td>
<td>184–220</td>
<td>9</td>
<td>3</td>
<td>0.651</td>
<td>184–212</td>
<td>4</td>
<td>1</td>
<td>0.490</td>
<td>186–210</td>
</tr>
<tr>
<td>LMS18</td>
<td>11</td>
<td>4</td>
<td>0.753</td>
<td>216–243</td>
<td>18</td>
<td>8</td>
<td>0.825</td>
<td>208–250</td>
<td>5</td>
<td>0</td>
<td>0.745</td>
<td>208–220</td>
</tr>
<tr>
<td>LMS28</td>
<td>5</td>
<td>3</td>
<td>0.694</td>
<td>291–299</td>
<td>3</td>
<td>1</td>
<td>0.202</td>
<td>289–293</td>
<td>1</td>
<td>0</td>
<td>0.000</td>
<td>291</td>
</tr>
<tr>
<td>LMS39</td>
<td>10</td>
<td>4</td>
<td>0.827</td>
<td>303–321</td>
<td>24</td>
<td>14</td>
<td>0.879</td>
<td>307–341</td>
<td>7</td>
<td>0</td>
<td>0.642</td>
<td>313–333</td>
</tr>
<tr>
<td>LMS45</td>
<td>4</td>
<td>0</td>
<td>0.193</td>
<td>295–307</td>
<td>11</td>
<td>7</td>
<td>0.595</td>
<td>295–328</td>
<td>4</td>
<td>0</td>
<td>0.619</td>
<td>298–307</td>
</tr>
<tr>
<td>LMS47</td>
<td>6</td>
<td>1</td>
<td>0.688</td>
<td>310–328</td>
<td>7</td>
<td>1</td>
<td>0.589</td>
<td>307–325</td>
<td>7</td>
<td>1</td>
<td>0.629</td>
<td>301–325</td>
</tr>
<tr>
<td>LMS53</td>
<td>2</td>
<td>0</td>
<td>0.301</td>
<td>175–178</td>
<td>2</td>
<td>0</td>
<td>0.011</td>
<td>175–178</td>
<td>1</td>
<td>0</td>
<td>0.000</td>
<td>175</td>
</tr>
<tr>
<td>LMS58</td>
<td>5</td>
<td>2</td>
<td>0.698</td>
<td>239–254</td>
<td>4</td>
<td>1</td>
<td>0.131</td>
<td>236–251</td>
<td>2</td>
<td>0</td>
<td>0.449</td>
<td>245–248</td>
</tr>
</tbody>
</table>

N\(_a\), number of alleles; N\(_p\), number of private alleles; H\(_e\) expected heterozygosity.
initially to believe that Chinese plant has ever been classified into section/series *Heterolespedeza*, members of which generally bear flattened buds and distichously arranged bud scales (incl. *L. buergeri* and *L. dunnii*). Among those that feature terete buds and spirally arranged bud scales, *L. thunbergii* (DC.) Nakai subsp. *formosa* (Vogel) H. Ohashi and *L. bicolor* Trucz. seem to be closely related to the Chinese plant. The former resembles Chinese plant because it has dense hairs on the leaflet surfaces as well as similarly shaped calyx lobes and leaflets. In fact, several specimens currently annotated as *L. maximoviczi* in the PE herbarium were originally identified as *L. formosa* (Vogel) Koehne, and have only recently been treated as synonyms of *L. thunbergii* subsp. *formosa* (Ohashi et al., 2009). However, *L. thunbergii* and its intra taxa usually produce longer flowers (ca. 14 mm long) than those of the Chinese plant, and the standard claw of *L. thunbergii* subsp. *formosa* (ca. 2.2 mm long) is shorter than those of the Chinese plant (ca. 2.7 mm long). The ratios of claw to lamina length in the wing (ca. 1:2.2) and keel petals (ca. 1:2.6) of *L. thunbergii* subsp. *formosa* are also different with those of Chinese plant (wing, ca. 1:1.3; keel ca. 1:1.4). *L. bicolor* is also similar to Chinese plant because of the ratio of claw and lamina lengths in the wing (ca. 1:1.5) and keel (ca. 1:1.5), and the fact that calyx lobed near middle part resembles with Chinese plant. However, *L. bicolor* bears standard petal attenuated base (Akiyama, 1988), whereas Chinese plant harbors standard petal with clawed base (e in Fig. 1B). In difference with Chinese plant, leaflets of *L. bicolor* usually are rounded to obtuse at apex, and glabrous. Although their apex of calyx lobes is acute, that of Chinese plant is acuminate. Consequently, Chinese plant is morphologically distinct from *L. thunbergii* subsp. *formosa* and *L. bicolor*.

**Taxonomic treatment**

*Lespedeza pseudomaximoviczi* D. P. Jin, B. Xu & B. H. Choi, sp. nov. (Figs. 1Ba–g, 4–6).—**TYPE**: CHINA. Zhejiang, Linan, Tianmu Mt., 18 Aug 2013, Bo Xu 2013-429 (Holotype:}

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**Fig. 4.** Chinese plant, named *Lespedeza pseudomaximoviczi*. **A.** Habit. **B.** Branch and leaves. **C.** Bark. **D.** Inflorescence.
Chinese name: Kuan-ye-hu-zhi-zi (宽叶胡枝子).

Shrub erect, 1–3 m tall, much branched at upper part. Branch terete, with sparsely adpressed hairs, light brown to brown, sometimes with black-colored dots; young branches greenish-brown. Leaves trifoliolate, petiolate, stipulate, pubescent; rachides 19.7–49.5 mm long; stipules, linear triangular to linear, brown to dark brown, 3.2–6.4 mm long; terminal leaflets gray-green, elliptic ovate to ovate, 39.1–68.1 mm long, 16.4–35.6 mm wide, acute to acuminate at apex, rounded or obtuse at base, upper surface pubescent, lower surface pubescent; petioles with adpressed hairs, 14.1–21.6 mm long; petiolules 1–2 mm long, swollen, adpressed or patent hairs. Inflorescence axillary, pseudo-raceme, usually one (rarely two or three) per one leaf, adpressed hair, 25.8–78.8 mm long, 14 to 36 flowered. Flowers 9.1–10.5 mm long, purple to pinkish-purple.

Bracteoles at base of calyx, ovate to oblong-ovate, pubescent, middle to above middle four-lobed; lobes subequal in length, acuminate at apex; lateral ones ovate; upper one broad ovate, two-cleft above middle. Standard 8.7–10.5 mm long, 4.1–5.5 mm wide, with auricles; lamina oblong to broad obovate, slightly emarginate at apex, purple inside, paler outside; claw 2.4–3.0 mm long. Wing petal 7.4–8.9 mm long, 1.7–2.2 mm wide; lamina oblong, 4.3–6.2 mm long; claw 3.1–4.0 mm long, with auricle. Keel petal 8.5–10.4 mm long, 1.9–2.5 mm wide; lamina slightly falcate, 4.3–6.2 mm long; claw 3.3–4.4 mm long. Legume broadly elliptic, ca. 10 mm long, ca. 5 mm wide, pilose, spine-tip.

Distribution: Anhui, Henan, Zhejiang (endemic species in China).

Flowering: (June–) July–August (–September).

Etymology: The specific epithet is derived from L. maximowiczii because of its rough resemblance.

Diagnosis: Lespedeza pseudomaximowiczii resembles L. maximowiczii in outward appearance, but differs in the petals...
with long claw and buds (terete, and spirally arranged scales), and in the pubescence on the upper surface of the leaflets. *Lespedeza thunbergii* subsp. *formosa*, is also similar to the *L. pseudomaximowiczii*, based on the shape and pubescence of the leaflets and shape of calyx lobes, but differs in flower size (ca. 14 mm long) and ratios of claw to lamina length in the petals (wing, ca. 1:2.2; keel, ca. 1:2.6). Even though wing and keel petals of *L. bicolor* shows similar ratios of claw and lamina lengths (wing, ca. 1:1.5; keel, ca. 1:1.5), its leaflets (glabrous on upper surface, and usually obtuse at apex) and standard petal with attenuated base are distinct from those of *L. pseudomaximowiczii*.


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**Conflict of Interest**

Authors declare that there are no conflicts of interest.

**Literature Cited**


